

## T cell receptor Zeta chain (TCR $\zeta$ ) expression in patients with idiopathic thrombocytopenic purpura

Dalia GA

Department of Clinical Pathology,  
Faculty of Medicine, Cairo University

### ABSTRACT

**Background:** T cell receptor zeta chain (TCR $\zeta$ ) plays a critical role in signal transduction via TCR. Defective signaling through TCR $\zeta$ -CD3 components may disrupt peripheral T cell tolerance to autoantigen, giving rise to a variety of autoimmune diseases. The aim of this work was to evaluate TCR $\zeta$  chain gene expression in patients with ITP, thereby to understand the functioning status of T cells and its possible contribution to disease outcome. **Methods:** SYBR Green based Real-time relative quantitative PCR (qRT-PCR) was used to evaluate TCR $\zeta$  gene expression in 49 ITP patients and 35 age- and sex-matched control subjects. **Results:** Patients in acute phase of ITP had a significantly lower TCR $\zeta$  gene expression compared to those in chronic phase and those of the control group; on the other hand TCR $\zeta$  gene expression was significantly lower in chronic ITP patients than in controls. Patients with active ITP had a significantly lower TCR $\zeta$  gene expression compared to those in remission and those of control group ( $P < 0.001$ ). TCR $\zeta$  gene expression was significantly lower in ITP patients in remission state in comparison to the control group. Treated ITP patients had a significantly higher TCR $\zeta$  gene expression compared to untreated patients, however TCR $\zeta$  gene expression in the treated group was still significantly lower compared to controls ( $P < 0.001$ ). Complete responders had a significantly higher TCR $\zeta$  gene expression compared to non responders ( $P < 0.001$ ), while no significant difference was found between non-responders and partial responders as regards TCR $\zeta$  gene expression ( $P > 0.05$ ). The level of TCR $\zeta$  gene expression in complete responders was still significantly lower compared to controls ( $P < 0.001$ ). Sequential analyses for TCR $\zeta$  gene expression in 6 patients with stable disease revealed no significant difference in median TCR $\zeta$  gene expression values in the 3 consequents analyses ( $P > 0.05$ ). TCR $\zeta$  gene expression showed no significant correlation with any of the clinical or hematological data. **Conclusion:** From these data, it could be suggested that downregulation of TCR $\zeta$  may represent intrinsic defects of potential pathogenetic significance for ITP. Therapy targeted to normalization of TCR $\zeta$  expression may represent a new strategy for treatment of ITP patients after being validated in clinical trials.

**Key words:** TCR $\zeta$ , qRT-PCR, ITP.

### INTRODUCTION

Idiopathic/immune thrombocytopenic purpura (ITP) is an autoimmune disorder in which anti-platelet antibodies induce platelets

destruction because of an imbalanced immune response. Although anti-platelet auto-antibody production is B-cell mediated, however it requires the presence of autoreactive T cells that are normally suppressed by the

regulatory T cells for immune tolerance<sup>1</sup>. In ITP a potentiated autoreactive T cells action secondary to diminished regulatory T cell numbers and/or function, is manifested suggesting loss of peripheral tolerance<sup>2,3</sup>.

The T cell receptor zeta chain (TCR $\zeta$ ) is a stable constituent of the TCR, it is present only in T- and natural killer-cells. TCR $\zeta$  chain, which also known as CD247, plays a critical role in the T-cell differentiation process and the T-cell effector function. Phosphorylation of TCR $\zeta$  chain is one of the earliest and key events in the T-cell signal transduction through the TCR-CD3 complex<sup>4</sup>. Even anergy or tolerance induction results from an active and selective signaling process involving phospho- $\zeta$  chain via the TCR<sup>5,6</sup>. Defective signaling through TCR $\zeta$ -CD3 components may disrupt peripheral T cell tolerance to autoantigen, giving rise to a variety of autoimmune diseases<sup>7</sup>.

Previous studies demonstrated a downregulated expression of the TCR $\zeta$  chain in several autoimmune diseases; systemic lupus erythematosus<sup>8</sup> (SLE), rheumatoid arthritis<sup>9</sup>, Type 1 diabetes mellitus<sup>10</sup> and ITP<sup>11</sup>.

Thus, detailed characterization of the expression of the TCR $\zeta$  chain may provide a key to understanding the molecular basis of T cell dysfunction in ITP patients.

#### **Aim of the work:**

The aim of this work was to evaluate TCR $\zeta$  chain gene expression in patients with ITP, thereby to understand the functioning status of T

cells and its possible contribution to disease outcome.

## **SUBJECTS & METHODS**

The present study was conducted on 49 patients with ITP as well as 35 age- and sex-matched control volunteers, after informed consent was obtained from each participant. These patients attended Kasr Al Aini Hospital, Cairo University, to be diagnosed in the Clinical Pathology Department between August 2010 and September 2011. ITP was diagnosed in accordance with the guidelines of the American Society of Hematology<sup>12</sup>. Thrombocytopenia was defined as platelet count  $<100 \times 10^9/L$ . The mean age of patients under study was  $43.4 \pm 17.8$  years. Among the 49 patients, 34 were females (69%) and 15 were males (31%). The clinical and laboratory data of the patients group are summarized in table 1.

Real-time relative quantitative PCR (qRT-PCR) with SYBR Green I technique was used to examine TCR $\zeta$  gene expression level in ITP patients and control subjects, beta 2 microglobulin ( $\beta_2M$ ) gene was used as an endogenous reference. RNA extraction from whole blood samples was done using QIAamp RNA Blood Mini Kit, (QIAGEN, Germany, catalog number 52304). Conversion of the extracted RNA to complementary DNA (cDNA) was done using RevertAid<sup>TM</sup> First Strand cDNA Synthesis Kit (Fermentas, USA, #K1622) according to the manufacturer's instructions. QRT-PCR was performed for TCR $\zeta$  and  $\beta_2M$  on Applied Biosystems 7300

التعليق [01]: Not recent

thermocycler (Qiagen, CA, USA) using SYBR<sup>®</sup> Green PCR Master Mix, (Applied Biosystems, Warrington, UK, Catalog Number 4309155) as previously described<sup>11</sup>. The 25  $\mu$ l reaction mixture contained 12.5  $\mu$ l SYBR Green qPCR Mix (2X), 1  $\mu$ l DNA, 1  $\mu$ l of each primer pair. The primers used for amplification were;  $\beta_2$ M internal control forward: 5'-TACTGAAATTCACCCAC-3', reverse: 5'-CATCCAATCAAATGCGGCA-3'. TCR $\zeta$  mRNA forward: 5'-GCCAGAACCAGCTCTATAAC-3', reverse: 5'-TAGGCCTCCGCCATCTT ATC-3'<sup>11</sup>. After the initial denaturation at 95°C for 2 min, 45 cycles consisting of 95°C for 15 s, 60°C for 1 min and 82°C for 1s were performed. The relative mRNA expression level of TCR $\zeta$  gene in each sample was calculated using the  $\Delta\Delta$  cycle time (Ct) method, where the target PCR Ct value, that is the cycle number at which emitted fluorescence exceeds the threshold line, is normalized by subtracting the  $\beta_2$ M Ct value from the target PCR Ct.

#### Statistical analysis:

Data were statistically described in terms of mean  $\pm$  standard deviation ( $\pm$  SD), median and range, or frequencies (number of cases) and percentages when appropriate. Comparison of numerical variables between the study groups was done using Mann Whitney U test for independent samples when comparing 2 groups and Kruskal Wallis test with posthoc multiple 2-group comparisons when comparing more than 2 groups. For comparing categorical data, Chi square ( $\chi^2$ ) test was performed.

Correlation between various variables was done using Spearman rank correlation equation for non-normal variables. P values less than 0.05 was considered statistically significant. All statistical calculations were done using computer programs SPSS (Statistical Package for the Social Science; SPSS Inc., Chicago, IL, USA) version 15 for Microsoft Windows.

## RESULTS

TCR $\zeta$  gene expression level in ITP patients ranged from 0.0002 to 18.406, with a median of 0.498, while in the control group, it ranged from 4.112 to 15.612, with a median of 6.308. TCR $\zeta$  gene expression level was significantly lower in ITP patients compared to the control group ( $P < 0.001$ ) (table 1, figure 1).

TCR $\zeta$  gene expression level ranged from 0.0002 to 18.150, with a median of 0.648 in ITP male patients and from 0.0002 to 18.406, with a median of 0.363 in ITP female patients, with no statistically significant difference ( $P > 0.05$ ) (table 1).

The clinical severity at the time of sampling was assessed by spontaneous bleeding, the bleeder group had a median TCR $\zeta$  gene expression of 0.368 (range of 0.0001-9.893), while the non-bleeder group had a median TCR $\zeta$  gene expression of 0.497 (range of 0.0001-18.406). Comparing bleeders and non bleeders as regards TCR $\zeta$  gene expression revealed no significant difference ( $P > 0.05$ ) (table 1).

According to the guidelines of the British Society for Hematology<sup>13</sup>, 30 (61%) of our patients were classified

**التعليق [02]:** Mention the concentrations of the DNA and primers

**التعليق [03]:** Be sure of the degree and the time

as acute ITP patients (disease duration < 6 months), and 19 (39%) were classified as chronic ITP patients, this classification was done at the time of sampling. Median TCR $\zeta$  gene expression was 0.200 (range of 0.0002-18.406) and 0.742 (range of 0.0002-7.378) in the acute and chronic ITP groups respectively. Patients in acute phase of ITP had a significantly lower TCR $\zeta$  gene expression compared to those in chronic phase and those of the control group, on the other hand TCR $\zeta$  gene expression was significantly lower in chronic ITP patients than in controls ( $P < 0.001$ ) (table 1, figure 1).

Defining the disease state at the time of sampling according to ZHOU et al. (2009)<sup>11</sup>, 39 out of 49 patients (79.5%) were in activity (platelet count  $< 100 \times 10^9/L$ ), and 10 out of 49 patients (20.5%) were in remission (platelet count  $\geq 100 \times 10^9/L$ ). Patients with active ITP had a significantly lower TCR $\zeta$  gene expression (median 0.188, range 0.0001-18.405) compared to those in remission (median 0.916, range 0.130-7.378) ( $P < 0.001$ ), and those of control group ( $P < 0.001$ ) (table 1, figure 1). On the other hand, TCR $\zeta$  gene expression was significantly lower in ITP patients in remission state in comparison to the control group ( $P < 0.001$ ) (table 1, figure 1).

At the time of sampling, 37 of the included ITP patients (75.5%) were under steroid therapy, among those steroid-treated patients 9 (18%) were receiving combination of steroids and Immuran. Treated ITP patients had a significantly higher TCR $\zeta$  gene expression (median 0.569, range 0.0002-9.894) compared to untreated

patients (median 0.165, range 0.0001-18.405) ( $P < 0.001$ ), however TCR $\zeta$  gene expression in the treated group was still significantly lower compared to controls ( $P < 0.001$ ) (table 1, figure 1).

Clinical and laboratory response were evaluated in 39 of the included patients (79.5%) who were available for a short period of follow up (3-6 months). They were further classified based on their platelet counts according to Francesco et al. (2009)<sup>14</sup> into complete responders (platelet count  $\geq 100 \times 10^9/L$ ), partial responders (platelet count between 30 and  $100 \times 10^9/L$ , and at least doubling of the baseline count) and non responders (platelet count lower than  $30 \times 10^9/L$  or less than doubling of the baseline count). The definition of response required concurrent resolution of bleeding symptoms. Corticosteroid- or other treatment-dependent patients were considered non-responders. Complete responders, partial responders and non responders, had a range TCR $\zeta$  gene expression of: 0.001-13.792, median of 0.823, 0.001-18.406, median of 0.138 and 0.001-1.469, median of 0.009 respectively. Complete responders had a significantly higher TCR $\zeta$  gene expression compared to non responders and to partial responders ( $P < 0.001$ ), while no significant difference was found between non responders and partial responders as regards TCR $\zeta$  gene expression ( $P > 0.05$ ). The level of TCR $\zeta$  gene expression in complete responders was still significantly lower compared to controls ( $P < 0.001$ ) (table 1, figure 1).

In order to examine for variability in the expression level of TCR $\zeta$  gene, sequential analyses for TCR $\zeta$  gene expression at two time points (2 months interval) after the base line expression at first sampling, were done for 6 chronic ITP patients selected according to the stability in the clinical course and the platelets count (<10% variation from sampling time count). No significant differences were found between the median expression levels of TCR $\zeta$  gene in the

1<sup>st</sup> (0.523, range of 0.0002-6.325), 2<sup>nd</sup> (0.498, range of 0.001-5.807) and 3<sup>rd</sup> (0.509, range of 0.008-4.956) analyses respectively (P>0.05) (table 1, figure 1).

TCR $\zeta$  gene expression showed no significant correlation with any of the clinical (patient age and disease duration) or hematological data (hemoglobin concentration, total leucocytes count and platelets count at sampling time and at follow up as well).

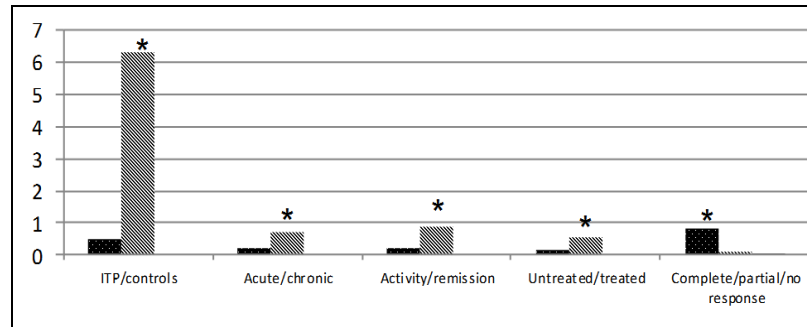
**Table 1:** Descriptive data of ITP patients, and a comparison of TCR $\zeta$  expression level in different groups of the studied ITP patients.

Variable	Value (Number,%)/(Mean $\pm$ SD)	TCR $\zeta$ expression Median, range	P-value
• ITP patients	49	0.498 (0.0002- 8.406)	<0.001
• Controls	35	6.308 (4.112 to 15.612)	
• Male	15 (31)	0.648 (0.0002-18.150)	> 0.05
• Female	34 (69)	0.363 (0.0002 - 18.406)	
Age in years	43.4 $\pm$ 17.8	-----	-----
Hemoglobin at sampling, g/dl	11.0 $\pm$ 2.2	-----	-----
TLC at sampling, x10 <sup>3</sup> / $\mu$ l	7.5 $\pm$ 4.4	-----	-----
Platelet at sampling, X 10 <sup>9</sup> /L	13 $\pm$ 60.3	-----	-----
Platelet at follow up, X 10 <sup>9</sup> /L	145 $\pm$ 113	-----	-----
• Acute	30 (61)	0.200 (0.0002-18.406)	< 0.001
• Chronic	19 (39)	0.742 (0.0002-7.378)	
• Activity	39 (79.5)	0.188 (0.0001-18.405)	< 0.001
• Remission	10 (20.5)	0.916 (0.130-7.378)	
• Bleeder	14 (29)	0.368 (0.0001-9.893)	> 0.05
• Non bleeder	35 (71)	0.497 (0.0001-18.406)	
No treatment	12(24.5)	0.165 (0.0001-18.405)	< 0.001
Treatment	37 (75.5)	0.569 (0.0002-9.894)	
• Steroids alone	28 (57.5)	-----	
• Steroids & Immuran	9 (18)	-----	
Total patients in follow up	39 (79.5)	-----	
• Complete response	24 (49)	0.823 (0.001-13.792)	< 0.001 <sup>a,b</sup>
• Partial response	8 (16.3)	0.138 (0.001-18.406)	> 0.05 <sup>a</sup>
• No response	7 (14.2)	0.009 (0.001-1.469)	-----
Sequential analysis:			
• 1 <sup>st</sup> analysis		0.523(0.0002-6.325)	
• 2 <sup>nd</sup> analysis	-----	0.498 (0.001-5.807)	> 0.05
• 3 <sup>rd</sup> analysis		0.509 (0.008-4.956)	

a: compared to no response

b: compared to partial response

التعليق [04]: What about the control sex??



**Figure1:** TCR $\zeta$  expression in controls as well as in different ITP patients groups.  
\* = P<0.05 (significant difference).

## DISCUSSION

T lymphocytes use a cell surface multisubunit structure, the TCR/CD3 complex, as an antigen-specific recognition site. The TCR  $\alpha\beta$  (or  $\gamma\delta$ ) chains are responsible for specific antigen binding, but they are unable to transmit any intracellular signals due to the very short cytoplasmic stretches they possess. Signal transduction is carried out by the T lineage-specific CD3 molecule, which is associated with the Ag-binding TCR chains, as well as with the TCR $\zeta$  chain dimer<sup>15,16</sup>.

One aspect of T cell physiology which is influenced significantly by the absence of the  $\zeta$  chain is the positive and negative selection of the immature thymus-dependent cells. Absence of the  $\zeta$  chain decreases both positive and negative selection. The decreased negative selection may enhance an autoimmune repertoire, and the decreased positive selection may be the basis for the decreased regulatory immune responses against autoreactivity<sup>17</sup>. Further support for the former hypothesis comes from a

study reporting an enhanced autoimmune potential for the T cells produced by  $\zeta$ -knockout mice<sup>18</sup>.

This study demonstrated that TCR $\zeta$  gene expression level was significantly lower in ITP patients compared to the control group. Reviewing literature, our study finding was in agreement with the only study that was found to analyze TCR $\zeta$  gene expression in ITP (18 chronic patients)<sup>11</sup>. In confirmation to the current study results, a significant decrease in TCR $\zeta$  gene expression was reported in other autoimmune diseases, including SLE<sup>19</sup>, RA<sup>20,21</sup> and Type 1 diabetes<sup>10</sup>. On the other hand, **Pang et al.**<sup>22</sup> reported that flow cytometric analysis demonstrated a significant decrease in the expression of TCR $\zeta$  in SLE, but in the other rheumatic diseases (RA, systemic sclerosis and primary Sjögren's syndrome), they noted a trend toward a decreased expression of TCR $\zeta$  but with no statistical significance.

No statistically significant difference was found in this study between males and females as regards TCR $\zeta$  gene expression levels. This

was in agreement to **Zhang et al.**<sup>11</sup> who reported no significant correlation between TCR $\zeta$  gene expression level and gender of ITP patients.

The current study reported that patients in acute phase of ITP had a significantly lower TCR $\zeta$  gene expression compared to those in chronic phase and those of the control group. On the other hand, TCR $\zeta$  gene expression was significantly lower in chronic ITP patients than in controls. Confirming to our findings, **Zhang et al.**<sup>11</sup> reported that chronic ITP patients had a significantly lower TCR $\zeta$  gene expression compared to healthy controls, it's worth noting that in their study none of the included ITP patients was in acute phase of the disease.

This study demonstrated that active ITP patient group had a significantly lower TCR $\zeta$  gene expression compared to those in remission and those of control group. On the other hand, TCR $\zeta$  gene expression was significantly lower in ITP patients in remission state in comparison to the control group. No similar analysis was found in literature for ITP patients. As regards other autoimmune diseases, **Pang et al.** (2002)<sup>22</sup> reported that one-third of the SLE patients exhibited a significant change in TCR  $\zeta$  expression at different levels of disease activity, while two-thirds of the patients showed a relatively stable low expression. No significant difference in the expression level of TCR $\zeta$  between the active and inactive patients was reported by **Liossis et al.**<sup>17</sup> in SLE patients. This discrepancy needs to be further studied with

consideration to different disease pathogenesis.

In the present study a significant up-regulation for TCR $\zeta$  was demonstrated in the steroid treated ITP patients versus untreated patients. In SLE, although **Pang et al.**<sup>22</sup> reported that three SLE patients in whom treatment could normalize the level of TCR  $\zeta$  expression, however their final statistical conclusion was that deficient TCR $\zeta$  chain expression is treatment unrelated. **Liossis et al.**<sup>17</sup> demonstrated that the decrease in TCR  $\zeta$  did not correlate with the dose of prednisolone.

In the current study among patients who were available for follow up, complete responders had a significantly higher TCR $\zeta$  gene expression compared to non responders and partial responders while no significant difference was found between the latter groups as regards TCR $\zeta$  gene expression.

Sequential analyses for TCR $\zeta$  gene expression revealed no significant differences between the median expression levels of TCR $\zeta$  gene in the 1<sup>st</sup>, 2<sup>nd</sup> and 3<sup>rd</sup> analyses. This was in agreement to **Pang et al.**<sup>22</sup> as they reported that two-thirds of the included SLE patients showed a relatively stable low expression of TCR $\zeta$  gene at two time points.

In this study, TCR $\zeta$  gene expression showed no significant correlation with any of the clinical or hematological data at sampling time and at follow up as well. In confirmation **Zhang et al.**<sup>11</sup> demonstrated no correlation between TCR $\zeta$  gene expression level with either age or gender of ITP patients.

Many theories have been postulated as how TCR  $\zeta$  defects lead to autoimmunity. In TCR  $\zeta$  chain knock-out mice, a shift from conventional thymic differentiated T cells to intestinal intraepithelial lymphocytes (IEL), which differentiate from the extrathymic environment, has been reported<sup>18</sup>. As it has been suggested that autoreactive T cell clones are more numerous in iIEL<sup>23</sup>, this could account for the autoimmune features in ITP with  $\zeta$  defects, as in  $\zeta$  knock-out mice.

The alternative hypothesis arises from demonstrations that autoreactive clones are not deleted by negative selection, but selected positively in thymic differentiation, and expanded in the periphery in TCR  $\zeta$  chain knock-out mice<sup>18,24,25</sup>, suggesting a role of the TCR $\zeta$  chain in negative selection in the thymus.

In addition, it has been proposed that the T cell anergy and altered receptor signaling in regulatory T cells (secondary to defective TCR $\zeta$  expression) may lead to autoimmunity by breaking the peripheral tolerance<sup>26</sup>.

Understanding the underlying defects within T lymphocytes is of critical importance not only for understanding disease pathophysiology, but also for identifying predictive biomarkers and better therapeutic targets.

Treatment of SLE patients with rapamycin, (also known as sirolimus), led to normalization of TCR $\zeta$  expression and calcium flux in T cells<sup>27</sup>. Lupus-prone mice treated with sirolimus showed reduced proteinuria, nephritis, and anti-dsDNA antibodies<sup>28</sup>. In a preliminary report,

seven of nine SLE patients refractory to conventional immunosuppression showed an improvement in disease activity scores after treatment with sirolimus<sup>29</sup>. Phase II trials of sirolimus in SLE and lupus nephritis are in progress.

In conclusion, from these data, it could be suggested that downregulation of TCR $\zeta$  may represent intrinsic defects of potential pathogenetic significance for ITP. Further experiments, for example showing the normalization of TCR-CD3 expression with genetic manipulation of TCR $\zeta$  to normal level, and concerning the detailed characterization of the downstream signaling molecules are required to understanding the molecular basis of TCR-CD3 dysfunction in ITP patients. Therapy targeted to normalization of TCR $\zeta$  expression may represent a new strategy for treatment of ITP patients after being validated in clinical trials.

## REFERENCES

1. **Craft J and Fatenejad S. (1997):** Self antigens and epitope spreading in systemic autoimmunity. *Arthritis Rheum.*, 40(8) : 1374-1382.
2. **Kuwana M, Kaburaki J, Kitasato H, Kato M, Kawai S, Kawakami Y, Ikeda Y. (2001):** Immunodominant epitopes on glycoprotein IIb-IIIa recognized by autoreactive T cells in patients with immune thrombocytopenic purpura. *Blood* 98(1): 130-139.
3. **Ogawara H, Handa H, Morita K, Hayakawa M, Kojima J, Amagai H, et al. (2003):** High



- Th1/Th2 ratio in patients with chronic idiopathic thrombocytopenic purpura. *Eur. J. Haematol.*, 71(4): 283-288.
4. **Eleftheriadis T, Antoniadi G, Liakopoulos V and Kortsaris A. (2006):** T-cell zeta chain expression, phosphorylation and degradation and their role in T-cell signal transduction and immune response regulation in health and disease. *Current Signal Transduction Therapy* 1(2): 191-208.
  5. **Lancaster SJ, Shaw AS, Rothbard JB. and Allen P M. (1994):** Partial T cell signaling: altered phospho-zeta and lack of zap70 recruitment in APL-induced T cell anergy. *Cell* 79: 913.
  6. **Madrenas J, Wange RL, Wang JL, Isakov N, Samelson LE and Germain RN. (1995):** Zeta phosphorylation without ZAP-70 activation induced by TCR antagonists or partial agonists. *Science* 267: 515-518.
  7. **Takeuchi T, Tsuzaka K, Pang M, Amano K, Koide J and Abe T (1998):** TCR z chain lacking exon 7 in two patients with systemic lupus erythematosus. *International Immunology* 10(7): 911-921.
  8. **Tsuzaka K, Setoyama Y, Yoshimoto K, Shiraishi K, Suzuki K, Abe T and Takeuchi T. (2005):** A splice variant of the TCR zeta mRNA lacking exon 7 leads to the down-regulation of TCR zeta, TCR/CD3 complex and IL-2 production in the systemic lupus erythematosus T cells. *Journal of Immunology* 174(6): 3518-3525.
  9. **Firestein GS (2004):** The T cell cometh: interplay between adaptive immunity and cytokine networks in rheumatoid arthritis. *The Journal of Clinical Investigation* 114(7): 471-474.
  10. **Nervi S, Atlan-Gepner C, Kahn-Perles B, Lecine P, Vialettes B, Imbert J and Naquet P (2000):** Specific deficiency of p56lck expression in T lymphocytes from type 1 diabetic patients. *Journal of Immunology* 165(10): 5874-5883.
  11. **Zhang XL, Li YQ, Chen SH, Yang LJ, CHEN S, WU XL, et al. (2009):** The feature of clonal expansion of TCR V $\beta$  repertoire, thymic recent output function and TCR  $\zeta$  chain expression in patients with immune thrombocytopenic purpura. *Int. J. Lab. Hematol.*, 31(6): 639-648.
  12. **Neunert C, Lim W, Crowther M, Cohen A, Solberg L, Jr. and Crowther MA. (2011):** The American Society of Hematology evidence-based practice guideline for immune thrombocytopenia. *Blood* 117(16):4190-4207.
  13. **British Society for Haematology. (2003):** Guidelines for the investigation and management of idiopathic thrombocytopenic purpura in adults, children and in pregnancy. Blackwell Publishing Ltd, *British Journal of Haematology* 120(4): 574-596.
  14. **Francesco R, Stasi R, Gernsheimer T, Blanchette, Kühne T, Ruggeri M, et al. (2009):** Standardization of

- terminology, definitions and outcome criteria immune thrombocytopenic purpura of adults and children: report from an international working group. *Blood* 113(11): 2386-2393.
15. **Weiss A and Littman DR. (1994):** Signal transduction by lymphocyte antigen receptors. *Cell* 76: 263–274.
  16. **Wange RL and Samelson LE. (1996):** Complex complexes: signaling at the TCR. *Immunity* 5:197–205.
  17. **Liou SC, Xuan Z D, Greg JD and Tsokos GC. (1998):** Altered Pattern of TCR/CD3–mediated Protein-tyrosyl Phosphorylation in T Cells from Patients with Systemic Lupus Erythematosus, Deficient Expression of the T Cell Receptor Zeta Chain. *The Journal of Clinical Investigation* 101(7): 1448–1457.
  18. **Yamazaki T, Arase H, Ono S, Ohno H, Watanabe H and Saito T (1997):** A shift from negative to positive selection of autoreactive T cells by the reduced level of TCR signal in TCR-transgenic CD3 zeta-deficient mice. *J. Immunol.*, 158(4): 1634–40.
  19. **Nambiar MP, Mitchell JP, Ceruti RP, Malloy MA, Tsokos GC. (2003):** Prevalence of T cell receptor zeta chain deficiency in systemic lupus erythematosus. *Lupus* 12(1) : 46–51.
  20. **Berg L, Rönnelid J, Klareskog L, Bucht A. (2000):** Down-regulation of the T cell receptor CD3 zeta chain in rheumatoid arthritis (RA) and its influence on T cell responsiveness. *Clin. Exp. Immunol.*, 120(1) : 174–182.
  21. **Romagnoli PD, Strahan M, Pelosi A, Cantagrel J, Meerwijk PV. (2001):** A potential role for protein tyrosine kinase p56<sup>lck</sup> in rheumatoid arthritis synovial fluid T lymphocyte hyporesponsiveness. *Int. Immunol.*, 13(3) : 305–312.
  22. **Pang, M, Setoyama Y, Tsuzaka K, Yoshimoto K, Amano K, Abe T, Takeuchi T. (2002):** Defective expression and tyrosine phosphorylation of the T cell receptor zeta chain in peripheral blood T cells from systemic lupus erythematosus patients. *Clin. Exp. Immunol.*, 129(1): 160–168.
  23. **Poussier P, Edouard P, Lee C, Binnie M, Julius M. (1992):** Thymus-independent development and negative selection of T cells expressing T cell receptor alpha/beta in the intestinal epithelium: evidence for distinct circulation patterns of gut- and thymus-derived T lymphocytes. *J. Exp. Med.*, 176(1) : 187–99.
  24. **Shores EW, Tran T, Grinberg A, Sommers CL, Shen H, Love PE. (1997):** Role of the multiple T cell receptor (TCR)-zeta chain signaling motifs in selection of the T cell repertoire. *J. Exp. Med.*, 185(5) : 893–900.
  25. **Shores EW, Ono M, Kawabe T, Sommers CL, Tran T, Lui Ket al. (1998):** T cell development in mice lacking all T cell receptor zeta family members (zeta, eta, and Fc epsilon RI gamma) *J. Exp. Med.*, 187(7) : 1093–101.

26. Saloijn KV, Zhang J, Madrenas J, Delovitch TL. (1998): T-cell anergy and altered T-cell receptor signaling: effects on autoimmune disease. *Immunol. Today*. 19(10) : 468–473.
27. Fernandez DR, Telarico T, Bonilla E, Li Q, Banerjee S, Middleton FA et al. (2009): Activation of mammalian target of rapamycin controls the loss of TCRzeta in lupus T cells through HRES-1/Rab4-regulated lysosomal degradation. *J. Immunol.*, 182(4) : 2063–2073.
28. Lui SL, Yung S, Tsang R, Zhang F, Chan KW, Tam S, Chan TM. (2008): Rapamycin prevents the development of nephritis in lupus-prone NZB/W F1 mice. *Lupus* 17(4): 305–313.
29. Fernandez D, Bonilla E, Mirza N, Niland B, Perl A (2006): Rapamycin reduces disease activity and normalizes T cell activation-induced calcium fluxing in patients with systemic lupus erythematosus. *Arthritis Rheum.*, 54(9): 2983–2988.

## التعبير عن سلسلة زيتا في مستقبلات الخلايا الليمفاوية تي في مرضى نقص الصفائح الفرغرى غير المسبب

داليا جميل امين

قسم الباثولوجيا الأكلينيكية - كلية الطب - جامعة القاهرة

سلسلة زيتا من مستقبل الخلايا الليمفاوية تي (TCR $\zeta$ ) تلعب دورا حاسما في نقل الإشارات عبر مستقبل الخلايا تي. تعطيل نقل الإشارات عبر TCR $\zeta$ -CD3 قد يؤدي الى فقد التواؤم الذاتي مما يؤدي إلى مجموعة متنوعة من أمراض المناعة الذاتية. يهدف هذا العمل الى تقييم التعبير عن TCR $\zeta$  كمحاولة لفهم الحالة الوظيفية للخلايا T في مرضى فرغرية نقص الصفائح مجهول السبب ودراسة إمكانية وجود علاقة بين مستوى التعبير عن TCR $\zeta$  ومسار المرض. تم تقييم التعبير عن TCR $\zeta$  باستخدام سلسلة البوليميراز الكمي في الوقت الفعلي بتحليل رد الفعل (PCR-qRT)، في 49 من المرضى الذين يعانون من أي تي بي، فضلا عن خمسة وثلاثون من الأصحاء كمجموعة ضابطة مطابقة في العمر والجنس. اسفرت الدراسة عن وجود انخفاض مستوى التعبير عن TCR $\zeta$  في مرضى ITP مقارنة بمجموعة الأصحاء. لم يظهر فرق ذا دلالة احصائية بين النساء والرجال في التعبير عن TCR $\zeta$ . كان مستوى التعبير عن TCR $\zeta$  أقل في مرضى ITP النشطة مقارنة مع الموجودة في هدأة والمجموعة الضابطة بينما كان مستوى التعبير عن TCR $\zeta$  أقل في مجموعة ITP الهدأة عن المجموعة الضابطة. كان مستوى التعبير عن TCR $\zeta$  أقل في مجموعة ITP الحاد بالمقارنة مع المرضى الذين يعانون من ITP المزمن والمجموعة الضابطة. برغم الارتفاع ذا الدلالة الاحصائية في مستوى التعبير عن TCR $\zeta$  بعد العلاج عن ما قبله الا انه لم يصل الى مستوى التعبير في المجموعة الضابطة. في مرضى المتابعة كان مستوى التعبير عن TCR $\zeta$  أعلى مرضى ITP المستجيبين مقارنة مع الغير مستجيبين ومرضى الإستجابة الجزئية بينما لم يختلف مستواه في المجموعتين الأخيرتين. التحليل التتابعى لمستوى التعبير عن TCR $\zeta$  في 6 من المرضى مستقرى الحالة لم يسفر عن اختلاف احصائى. لم يثبت وجود علاقة بين مستوى التعبير عن TCR $\zeta$  وأى من البيانات الاكلينيكية او المعملية للمرضى. الدراسة الحالية تؤكد التقارير السابقة التى تشير إلى انخفاض التعبير عن TCR $\zeta$  في المرضى الذين يعانون من مرض ITP النشط. وكانت الإستجابة للعلاج والهدأة فى المرض مصاحبه للزيادة في التعبير عن TCR $\zeta$  معبرة عن اتصاله بشدة المرض. المعالجة التي تستهدف الحث على التعبير عن TCR $\zeta$  تحمل أملا جديدا لمرضى فرغرية نقص الصفائح مجهولة السبب.