EVALUATION OF ACUTE TOXICITY AND TERATOGENIC EFFECTS OF SOME PLANT GROWTH REGULATORS ON ALBINO RAT EMBRYO

El-sayyad H. I.*; M. H. Abo Egla**; M. M. Mortada*; Manal M. Ramadan* and M. E. Elbeeh *

* Fac. Sci., Mans. Univ.,**Plant Protection Res. Inst.,Ministry Of Agric., Egypt

ABSTRACT

Gibberellic acid(GA_3) and Indole acetic acid (IAA) is an endogenous plant growth regulator used world wide in agriculture; however, little is known about its biochemical and physiological effects on mammals. This study investigated possible toxic and teratogenic effects of GA_3 and IAA on the embryo of albino rats. Female Wistar rats were given daily 200 ppm / 0.2 ml saline from either GA_3 or IAA orally from the 1st day of pregnancy until the 14^{th} , 16^{th} , or after delivery. Toxicity was demonstrated by a significant increase in malondialdehyde level and a decrease in the antioxidant enzyme activities of catalase, superoxide dismutase, and glutathione peroxidase in the brain, spinal cord, eye, and liver of pups. A significant decline of glutathione content was also observed. The biochemical parameters were correlated histologically with an abnormal development of the external morphology and formation of the skeleton of either GA_3 or IAA-treated pups.

INTRODUCTION

Gibberellins are a group of plant growth regulators widely use to improve the yield of a wide variety of plants by increasing cell division, cell elongation that affect leaves as well as stems (Silverstone and Sun, 2000) and accelerate the growth of fruits and vegetables in the world (Arous *et al.*, 2001).

The World Health Organization (WHO, 1990) listed gibberellin-A₃ as a plant growth regulators related to pesticides.

Indole acetic acid is a plant hormone detected in human urine (Qureshi and Baig, 1993), blood plasma (as a metabolite of tryptophan) (Bertuzzi et al., 1997) and central nervous system (Hu and Dryhurst, 1997). It is also, found in cerebrospinal fluid and in several organs such as liver, kidney, lungs and brain (De Melo et al., 1998). IAA is predominantly formed as a result of the mono amin oxidase - mediated oxidative de amination of tryptamine, a putative neurotransmitter or neuromodulator in CNS (Artigas et al., 1983). The plasma levels of IAA and its metabolites were elevated in some human diseases such as insulin dependent diabetes mellitus (Rogerson et al., 1991), renal dysfunction and phenylketonuria (Qureshi and Baig, 1993). The combination of IAA and horseradish peroxidase (HRP) was found to cause cytotoxic toward mammalian cells (Folkes et al., 1999) and increase lipid peroxidation (Candeias et al., 1996; Folkes et al., 1999).

The toxic effects of these chemicals on animals are limited; therefore, this subject has attracted the interest of many researchers recently. Many

chemicals are currently used in agriculture, and PGRs are among those widely used. The amounts of these substances placed into the environment may soon exceed those of insecticides (Mickey, 1978). The effects of different PGRs on insects have been investigated (Visscher, 1983; De Man et al., 1991), but reports concerning vertebrates are very limited (El-Mofty and Sakr, 1988; Ustun et al., 1992; Ozmen et al., 1995; de Melo et al., 2004; Furukawa et al., 2004; Hsiao et al., 2004). In the literature, it is reported that fecundity, longevity, and egg viability have been changed in different insects by PGRs treatment (De Man et al., 1991). Furukawa et al. (2004) indicate that indole acetic acid (IAA) might induce the neuronal apoptosis in the S phase and lead to microencephaly.

Concerning its effect on apoptosis, Furukawa et al. (2004) that IAA induces neuronal apoptosis in S phase and leads to microencephaly. According to previous studies (Candeias et al., 1995; Celik and Tuluce, 2006; Tuluce and Celik, 2006; Celik et al., 2007). Gibberellic acid(GA₃) is the most commonly used PGRs in agriculture in many countries, including Tunisiain order to enhance fruit growth like date palm (Ben Abdallah et al., 2000) and some vegetables such as pepper(Arous et al., 2001) and olive (Chaari-Rkhis et al., 2006). GA₃ has also been shown to cause alarming toxicity to mammalian systems, particularly in the breast, lung (El-Mofty et al., 1994), kidney and liver (Ustun et al., 1992) of adult mice. In addition, GA₃ administration by gavage for 22 months induces carcinogenic effects in adult Swiss Albino mice (El-Mofty et al., 1994). According to Ozmen et al. (1995) treatment with GA₃ affects sexual differentiation and some physical parameters in laboratory mice. Recent reports indicate that this PGRs may induce oxidative stress, leading to the generation of free radicals and causing cells damage in many organs, including the heart, kidney, stomach and spleen of adult rats (Celik and Tuluce, 2006) and the liver of GA3 treated suckling rats (Troudi et al., 2010).

MATERIALS AND METHODS

All experiments were conducted in accordance with the national laws for the use of animals in research and approved by the local ethical committee.

Chemicals:

Two applied plant hormones were used in the present work; one belong to Gibberellin group (Gibberellic acid) and the other belong to auxin (indoleacetic acid) were purchased from Sigma Chemical Company (St. Louis, MO 6, USA). Both compounds`were dissolved in tap water at doses of 200 ppm / 0.2 ml saline for GA_3 and IAA.

Gibberellic acid

Indole acetic acid:

Experimental work:

Seventy-two fertile male and virgin female rats weighing approximately 125g body weight were obtained from Hellwan Breeding Farm, Ministry of Health at ratio of 1:3 respectively and used for experimentation. Vaginal smears were carried out to give a precise determination of the onset of gestation.

The rats were arranged in to three groups, each was composed of 18 individuals as follows:

- 1- Control pregnant: the animals received basal diet and tab water.
- 2– Gibberellic acid (GA₃)-treated pregnant: Each individual received oral doses of 200 ppm / 0.2 ml saline for 40 days every other day and from the 1st to 14th, 16th, or delivery day of gestation.
- 3– Indoleacetic acid (IAA)-treated pregnant: Each individual received oral doses of 200 ppm/ 0.2 ml saline for 40 days every other day and from the 1st to 14th, 16th, or delivery day of gestation.

RESULTS

From table (1), treating pregnant with either gibberellic or indole acetic acid on pregnant mothers revealed 5/25 of pregnant mothers failed to complete pregnancy especially of those subjected to indole acetic aci treatment. Experimental group receiving gibberellic acid -treatment showed marked decrease of the number of delivered pups. There was a marked reduction of both body weights and crown-rump size of prenatal embryos at 14 & 16 days old as well as pups maternally subjected to plant growth hormone-treatment especially of those subjected to indole acetic acid-treatment.

Examining the gross morphology of pups of experimental treated groups revealed the presence of different pattern of congenital malformations. Superficial haematomas and fore limb deformation were detected in delivered newborn of studied experimental groups. Highest incidence was recorded in experimental epileptic group receiving indole acetic acid—treatment (Table (2-5), Figs. 1-9).

Table (1): Effect of experimentally either Gibberellic acid (GA₃) or Indole Acetic acid (IAA) treated female albino rats during pregnancy on pregnant mothers and their fetuses and pups.

	Control	(GA ₃)	(IAA)
Total number of mothers	20 (100%)	25 (100%)	25 (100%)
Total number and percentage of aborted mothers	0 (0%)	5 (20%)	5 (20%)
Total number and percentage of pregnant	20 (100%)	20 (80%)	20 (80%)
Total number of fetuses and newborn	120 (100%)	60 (100%)	750(100%)
% of numerical reduction of fetuses & newly born from control	0 (0%)	(50%)	(62.5%)
Total number (% of fetal mortality)	0 (0%)	17 (28.4%)	21 (28%)
Total number & % of alive fetuses & newly born	120 (100%)	43 (71.6%)	54 (72%)

Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

Table (2): Body weight (gm) and size (ml³.) of fetuses (14&16 -days) and pups treated with either (GA₃) or (IAA).

Pregnan cy Day	14 day prenatal			16	day prena	atal	pups			
	С	C GA ₃ IAA		C GA ₃ IAA		GA ₃ IAA C GA ₃ IAA		С	GA₃	IAA
weight	4.01±	3.60±	3.30±	4.50±	4.10±	3.90±	5.30±	4.59±	4.67±	
weight	0.04	0.03*	0.01*	0.06	0.04*	0.06*	0.01	0.04*	0.04*	
size	3.9±	2.4±	2.8±	5.6±	4.1±	4.5±	15.8±	9.5±	9.2±	
Size	0.07	0.051*	0.056*	0.051	0.088*	0.052*	0.094	0.13*	0.121*	

Each result represents the mean \pm SD of 10 replicates.

^{*} Significant at p < 0.05. Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

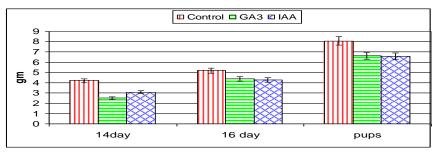


Fig. 1: Body weight (gm.) of fetuses (14, 16) and pups treated with either (GA₃) or (IAA).

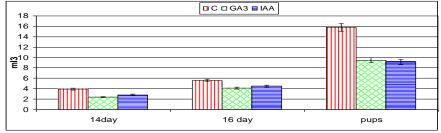


Fig. 2: Body size (ml3.) of fetuses (14&16days) and pups treated with either (GA_3) or (IAA).

Table (3): Crown-rump length (cm.) of fetuses (14&16) days and pups treated with either (GA₃) or (IAA).

14 day prenatal			16	day prena	21 day			
С	GA₃	IAA	A C GA ₃ IAA C GA ₃				IAA	
1.99±0.08	1.76±	1.7±	2.55±	2.03±	2.02±	4.1±	3.74±	3.84±
1.33±0.00	0.063*	0.053*	0.223	0.144*	0.108*	0.047	0.054*	0.107*

Each result represents the mean ± SD of 10 replicates.

^{*} Significant at p < 0.05.Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

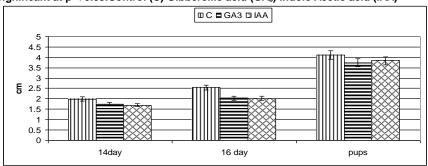


Fig.3: crown-rump length (cm.) of 14&16 days fetuses and pups treated with either gibberellic (GA₃) or indole acetic acids (IAA).

Table (4): Incidence of gross morphological and skeletal abnormalities of 14&16 days fetuses and pups treated with either (GA₃) or (IAA).

	Control	GA ₃	IAA
Total number & % of alive fetuses & newly born	120 (1000%)	43 (71.6%)	54 (72%)
Superficial haematomas	0 (0%)	9 (20.93%)	10 (18.51%)
Abnormal fore limb			
-Unilateral	0 (0%)	12 (27.9%)	15 (27.8%)
-Bilateral	0 (0%)	20(46.51%)	25 (46.29%)
Abnormal hind limb			
-Unilateral	0 (0%)	9 (20.93%)	7 (12.96%)
-Bilateral	0 (0%)	20 (46.51%)	30(55.55%)
Incomplete ossification of sternum	0 (0%)	8(18.60%)	6(11.11%)
Kyphotic body	0 (0%)	10(23.25%)	5(9.25%)
Kinky tail	0 (0%)	16(37.20%)	14(29.92%)
Missing of caudal vertebrae	0 (0%)	11(25.58%)	13(24.07%)

Table (5): Head length & width (cm.) of 14&16 days fetuses and pups treated with either (GA3) or (IAA).

				<u>, , , , , , , , , , , , , , , , , , , </u>	_ ,					
Pregnancy Day	14	day pren	atal	16	day pren	atal	21 day			
	С	GA₃	IAA	С	GA₃	IAA	С	GA₃	IAA	
length	0.82±	0.65±	0.59±	0.9±	0.80±	0.83±	1.08±	0.93±	0.91±	
lengui	0.032	0.033*	0.022*	0.017	0.012*	0.021*	0.01	0.034*	0.039*	
width	0.49±	0.40±	0.42±	0.57±	0.47±	0.49±	0.71±	0.59±	0.64±	
widin	0.036	0.025*	0.014*	0.009	0.022*	0.031*	0.009	0.038*	0.024*	

Each result represents the mean ± SD of 10 replicates.

^{*} Significant at p < 0.05.Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

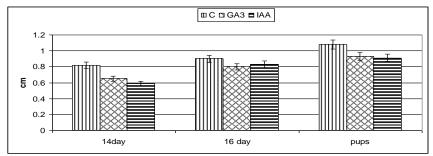


Fig.4: Head length (cm.) of 14&16 days fetuses and pups. Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

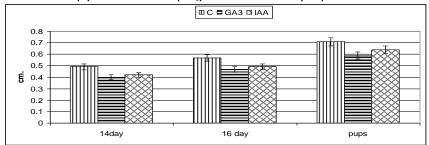


Fig.5: Head width (cm.) of 14&16 days fetuses and pups. Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

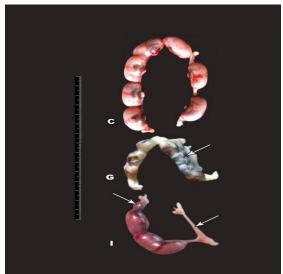


Fig. 6 (C-G): photomacrographs of uterus of 16 days of gestation. C. control uterus showing symmetric distribution of implantation. G. Mother's uterus treated with (GA₃) & I. Mother's uterus treated with (IAA) showing resorption of fetuses and asymmetric distribution of implantation.



Fig. 7 (C-I2): Photomacrographs of lateral view gross morphology of 14, 16-days old fetuses and pups. C. Control 14-days old fetuses. G. 14-day old fetuse maternally treated with (GA₃) showing deformation of limbs and superficial haematomes. I. 14 days old fetuses maternally (IAA) showing deformation of limbs. C1. Control 16-days old fetuses. G1. & I1. 16-day old fetuses showing marked deformation in size, fore and hid limb, non demarkation of trunk region and superficial haematomes. C2. Control pups. G2. & I2. Pups showing marked deformation in size, deformation of fore and hid limb, exencephaly and superficial haematomes.

3. Effects of ossification of skeleton of delivered newborn:

Tables (6-7) revealed that pups maternally-treated with either gibberellic or indole acetic acid showed a marked retardation of ossification centers in axial and appendicular regions. Abscence of ossification center restrict mainly in interparietal, incus, malleus, stapes, and tympanic bone, exoccipital and supraoccipital regions. Abscence of ossification center were noted also in metacarpales, distal phalanx of fore limb, metatarsals, distal phalanx of hind limb and sternum. The highest incidence of missing ossification center was detected in prenatal embryos and pups maternaly treated with indole acetic acid (Figs.13).

The ossified length of mandibular and fore-(Humerus, radius & ulna) and hind limb parts (femur, tibia and fibula) were markedly reduced in prenatal embryos (14&16 days) as well as in pups maternally treated with either gibberellic or indole acetic acid-treatment (Tables 8-9, Figs.8 - 10).

Tables (10&11) illustrate calcium content in maternal serum and femur bone as well as in skeleton of prenatal embryos (14&16 days) and pups

maternally subjected to either gibberellic or indole acetic acid treatment. In case of mothers, treatment with either gibberellic or indole acetic acid led a decrease of maternal serum and femur bone calcium. On the other hand, there was a marked reduction of total calcium content in prenatal embryos and pups maternally treated with either gibberellic or indole acetic acid (Figs. 11&12).

Table (7): Incidence of skull bone abnormalities of pups treated with either (GA₃) & (IAA).

Group	Control	GA3	IAA
Total number & % of alive fetuses & newly born	120 (1000%)	43 (71.6%)	54 (72%)
Nasal	0 (0%)	4 (9.30%)	11 (20.37%)
Frontal	0 (0%)	10 (23.25%)	15 (27.78%)
Parietal	0 (0%)	6 (13.95%)	8 (14.81%)
Inter parietal	0 (0%)	12 (27.90%)	11 (20.37%)
Maxilla	0 (0%)	4 (9.30%)	2 (3.70%)
Zygomatic arch	0 (0%)	6 (13.95%)	2 (3.70%)
Hyoid Arch	0 (0%)	4 (9.30%)	4 (7.40%)
Squamosal	0 (0%)	6 (13.95%)	4 (7.40%)
Incus	0 (0%)	8 (18.60%)	2 (3.70%)
Malleus	0 (0%)	12 (27.90%)	8 (14.81%)
Stapes	0 (0%)	12 (27.90%)	11 (20.37%)
Tympanic ring	0 (0%)	4 (9.30%)	4 (7.40%)
Supraoccipital	0 (0%)	7 (16.27%)	6 (11.11%)

Table (8): Ossified length (mm.) of Mandibular and fore limb of 14&16 days fetuses and pups treated with either (GA₃) or (IAA).

Pregnancy Day	14 day prenatal			16 day prenatal			pups			
	С	GA₃	IAA	С	GA₃	IAA	С	GA₃	IAA	
Mandibular	4.2±	2.5±	3.1±	5.2±	4.4±	4.3±	8.09±	6.64±	6.57±	
Wandibular	0.02	0.01*	0.02*	0.011	0.01*	0.03*	0.1	0.46*	0.14*	
Humerus	4.7±	2.4±	1.68±	5.64±	4.1±	3.92±	6.45±	4.750±	5.9±	
numerus	0.05	0.06*	0.07*	0.05	0.1*	0.08*	0.03	0.08*	0.11*	
Radius	2.7±0.06	1.8±	1.7±	5.67±	3.8±	3.08±	9.03±	6.56±	7.08±	
Radius	2.7±0.06	0.04*	0.06*	0.07	0.06*	0.08*	0.09	0.07*	0.09*	
Illno	1.60±	1.21±	0.92±	4.08±	2.7±	2.8±	5.85±	3.14±	2.84±	
Ulna	0.03	0.04*	0.05*	0.05	0.07*	0.07*	0.04	0.10*	0.05*	

Each result represents the mean \pm SD of 10 replicates.

^{*} Significant at p < 0.05. Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

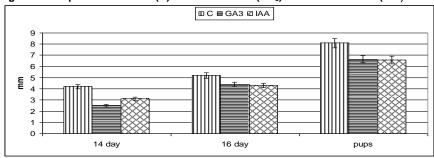
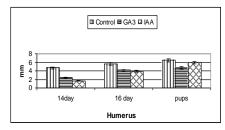
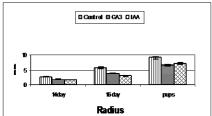


Fig.8: Ossified length (mm.) of Mandibular bone of 14&16 days fetuses and pups. Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

J. Plant Prot. and Path., Mansoura Univ., Vol. 3 (9), September, 2012





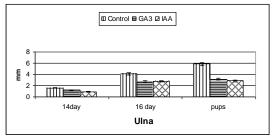


Fig.9: Ossified length (mm.) of fore limb of 14&16 days fetuses and pups. Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

Table (9): Ossified length (mm.) of hind limb of 14&16 days fetuses and pups.

Pregnancy Day	14 (14 day prenatal			16 day prenatal			pups			
	С	GA₃	IAA	С	GA₃	IAA	С	GA₃	IAA		
Femur	3.32±	1.44±	0.8±	4.84±	2.44±	2.8±	5.85±	3.14±	2.84±		
remui	0.03	0.06*	0.02*	0.05	0.04*	0.07*	0.04	0.10*	0.05*		
Tibia	2.96±	2.18±	1.48±	7.0±	4.1±	2.6±	6.25±	3.79±	3.32±		
Tibla	0.04	0.05*	0.05*	0.05	0.05*	0.05*	0.05	0.07*	0.16*		
Fibula	2.8±	1.9±	1.4±	4.72±	3.12±	2.72±	4.82±	2.67±	2.05±		
Fibula	0.05	0.05*	0.04*	0.04	0.24*	0.24*	0.03	0.10*	0.11*		

Each result represents the mean ± SD of 10 replicates.

* Significant at p < 0.05. Control (C) Gibberellic acid (GA $_3$) Indole Acetic acid (IAA)

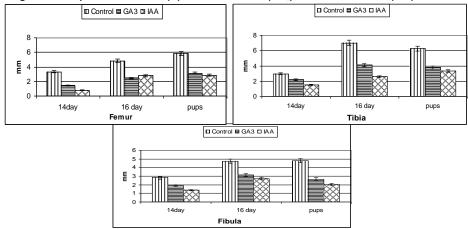


Fig.10: Ossified length (mm.) of hind limb of 14&16 days fetuses and pups. Control (C) Gibberellic acid (GA₃) Indole Acetic acid (IAA)

Table (10): Calcium content in Serum and femur bone of mothers intoxicated with either (GA3) or (IAA).

	С	GA3	IAA
Serum (µg / ml)	2.40 ± 0.10	2.07 ± 0.15*	1.70 ± 0.20*
Femur (g / g bone)	0.48 ± 0.01	0.40 ± 0.02*	0.39 ± 0.02*

Each result represents the mean ± SD of 10 replicates.* Significant at p < 0.05.

Table (11): Calcium content in the skeleton of 14&16 days fetuses and pups maternally treated with either (GA3) or (IAA).

	Pape III		1. 0 0.10 0.		(/ 01 (11 11	.,.		
14 day				16 day		pups			
С	GA3	IAA	С	C GA3 IAA			GA3	IAA	
0.13±0.01	0.10±0.02*	0.09±0.03*	0.15±0.01	0.12±0.01*	0.11±0.02*	0.19±0.01	0.16±0.02*	0.15±0.05*	

Each result represents the mean \pm SD of 10 replicates. * Significant at p < 0.05.

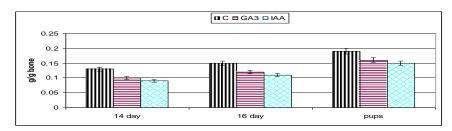


Fig. 11: Calcium content in the skeleton of 14&16 days fetuses and pups.

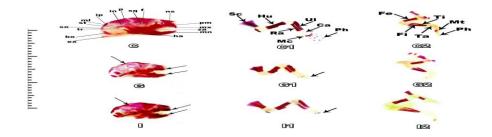


Fig.12 (C-I): Lateral view macrograph of skull, fore limb and hind limb of Alizarin red S preparation of pups intoxicated with applied plant growth hormones. C. Control skull bones. G. Pups skull maternal treated with (GA₃) Showing reduction of flat skull bones and absence of incisors. I. Pups skull maternal treated with (IAA) showing reduction of ossification center of parietal bones and absence of incisors. C1. Control pups fore limb. G1. Pups fore limb maternal treated with (GA₃). I1. Pups fore limb maternal treated with (IAA). Note reduction of ossified bones and lack ossification of phalanges. C2. Control pups hind limb. G2. &I2: Pups hind limb maternal treated with (GA₃) & (IAA). Note reduction of ossified bones and lack ossification of phalanges.

(Abbreviations: ns: Nasal, f: Frontal, p: Parietal, ip: Inter parietal, mx: Maxilla, za: Zygomatic arch, ha: Hyoid Arch, sq: Squamosal, in: Incus, ml: Malleus, st: Stapes, tr: Tympanic ring, so: Supraoccipital, ex: exoccipital, bo: basocipital, Sc: scapula, Hu: humerus, Ra: radius, Ul: ulna, Ca: carapace, Mc: metacarapace, ph: phalanges, Fe: femur, Ti: tibia, Fi: fibula, Ta: tarsas, Mt: metatarsas, ph: phalanges.)

DISCUSSION

Recently plant growth hormones are among those widely used chemicals in agriculture accelerated the growth of fruits and vegetables (Serrani *et al.*, 2007a&b). The amounts of its placed into the environment exceed those of the insecticides (Mickel, 1978). Although, the consumption of contaminated food products led these growth hormones to find their ways in our body, however, the toxicological aspects are very limited (El-Mofty and Sakr, 1988; Ustun *et al.*, 1992; Ozmen et al., 1995; de Melo *et al.*, 2004; Furukawa *et al.*, 2004; Hsiao *et al.*, 2004).

From the present findings, oral administration of either gibberellic or indole acetic acid at doses of 2·0 ppm for 40 days every other day and from the 1st to 14th day of gestation led to alterations in maternal hepatic tissues and femoral bone. Liver hepatitis was characterized by either cytoplasmic vacuolization of the hepatocytes with increased incidence of pyknotic nuclei. There was a marked increase of dissolution of hepatic cords with prominent dilated blood sinusoids and ill defined cell boundaries of hepatocytes. Numerous hypertrophied kupffer cells were detected in the sinusoidal wall. The blood vessels become either congested or hyalinized with apparent degeneration of their lining endothelium as well as had perivascular leukocytic infiltration. Drastic effects of maternal livers were reflected on the liver of their pups of almost identical histopathological lesions in the form of massive cell death characterized by massive necrosis of hepatocytes, distortion of blood sinusoids and abnormal congestion of blood vessels with apparent degeneration of their endothelial lining cells.

Similar findings were reported in pregnant rats previously received daily 200 ppm gibberellic acids in drinking water from the 14th day of pregnancy until 14 days post partum (Trodi *et al.*, 2010) as well as in rat received 25 ppm for 21 days (Sakr *et al.*, 2003) or 75 ppm GA₃ for either 50 days (Soliman *et al.*, 2010) or six weeks (Hussein et al., 2011).

The observed hepatitis of both mothers and their pups was supported by abnormal biochemical markers of liver functions including sera ALT & AST, billirubin, albumen, arginase and D-L- Fucosidae.

The present data revealed that pups and fetuses maternally treated with gibberellic or indole acetic acid exhibited marked reduction of body weight and crown-rump length. The rate of morphological abnormalities was markedly increased and characterized by kyphotic body, malformed fore and hind limb, kinky tail, reduced neck region and superficial haematomas. Similar findings were reported by Furukawa *et al.* (2004) who observed microencephaly and cleft palate in rat pups maternally treated with indole acetic acid.

The observed intrauterine growth retardation may be attributed to transplacental passage of the used plant growth hormones or their metabolites interfering with growth differentiation of developing fetuses.

In general, ossification of skull regions carries out by two mechanisms; endochondral and intramembranous ossification. During endochondral ossification, precursor cell condense in area destined to become bone and

acquire the general shape of the bone segments, thus providing the template for future skull elements (Hall and Miyake, 1995). This mechanism is prevalent leading to the formation of base and caudal parts of skull (chondrocranium). The rest of the skull, including cranial vault and maxillamandibular bones, is formed by intramembranous ossification, a process which involved the condensation of mesenchymal precursors and the transition to the differentiated bone cells, without an intermediate cartilaginous template (Hall and Miyake, 1992).

The observed findings revealed that pups maternally intoxicated with gibberelic or indole acetic acid exhibited marked inhibition of both kinds of axial and appendicular bones. Delayed ossification may be attributed to the impaired osteoblast maturation and function causing skull abnormalities. The observed results revealed that fetuses and pups of experimental treated groups exhibited marked reduction of ossified bones including mandible, humers, radius, ulna, femur, tibia, fibula, ilium and ischium.

The skeletal defects of fetuses and pups may be resulted from both pathological alterations of the maternal liver which intern reflected on liver of pup causing marked reduction of both protein and calcium (Bellomo and Orrenius, 1985; Vessey, 1996) and vitamin D synthesis (Kochupillai, 2008; Lehmann and Meurer, 2010).

The observed gross deformity and delayed ossification of bones may be attributed to the increased oxidative stress which influenced by DNA damage and intern interfere with cell replication leading to growth defects (Troudi *et al.*, 2011).

The authors finally concluded that to advise farmers to reduce application of plant growth hormones in their green house to reduce their impacts on health along run of life.

REFERENCES

- Arous S, Boussaid M, Marrakchi M (2001). Plant regeneration from zygotic embryo hypocotyls of Tunisian chilli (Capsicum annuum L.). J Appl Hortic.; 3:17–22.
- Arous S, Boussaid M, Marrakchi M (2001). Plant regeneration from zygotic embryo hypocotyls of Tunisian chilli (Capsicum annuum L.). J Appl Hortic.; 3:17–22.
- Artigas F, Martinez E, Tusell JM, Sunol C, Gelpi E (1983). On the metabolic origin of plasmatic indole-3-acetic acid in the rat. Biochem Pharmacol.; 32: 3251–54.
- Bellomo G, Orrenius S (1985). Altered thiol and calcium homeostasis in oxidative hepatocellular injury. Hepatology; 5: 876-882.
- Ben Abdallah A, Stiti K, Lepoivre P, Dujardin P (2000). Identification de cultivars de palmier dattier (Phoenix dactylifera L.) par l'amplification aleatoire d'AND (RAPD). Cah Agric.; 9: 103–7.
- Bertuzzi A, Mlngrone G, Gandolfi A, Greco AV, Ringoir S, Vanholder R (1997). Binding of indole-3-acetic acid to human serum albumin and competition with L-tryptophan Clinica Chimica Acta.; 265: 183-192.

- Candeias LP, Folkes LK, Porssa M, Parrick J, Wardman P (1995). Enhancement of lipid-peroxidation by indole-3-acetic acid and derivatives-substituent effects. Free Radic Res.; 23: 403–418.
- Candeias LP, Folkes LK, Wardman P (1996). Enhancement of peroxidase-induced lipid peroxidation by indole-3-acetic acid: effect of antioxidants. Redox Rep.; 2: 141–147.
- Celik I, Tuluce Y (2006). Effects of indole acetic acid and kinetin on lipid peroxidation and antioxidant defense in various tissues of rats. Pest Biochem Physiol.; 84: 49–54.
- Celik I, Turker M, Tulice Y (2007). Absisic acid and gibberellic acid cause increased lipid peroxidation and fluctuated antioxidant defense systems of various tissues in rats. J Hazard Mater; 148: 623-629.
- Chaari-Rkhis A, Maalej M, Ouled Messaoud S, Drira N (2006). Invitro vegetative growth and flowering of olive tree in response to GA3 treatment. Afr J Biotechnol.; 5: 2097–302.
- De Man W, De Loof A, Briers T, Huybrechts R (1991). Effect of abscisic acid on vitellogenesis in Sarcophaga bullata, Entomol Exp Appl.; 29: 259–267.
- De Melo MP, De Lima TM, Pithon-Curi TC, Curi R (2004). The mechanism of indole acetic acid cytotoxicity. Toxicol.; Lett. 14: 148(1–2): 103–111.
- De Melo MP, Pithon-Curi TC, Miyasaka CK, Palanch AC, Curi R (1998). Effect of indole acetic acid on oxygen metabolism in cultured rat neutrophils. Gen Pharmacol.; 31: 573–578.
- El-Mofty MM, Sakr SA (1988). Induction of neoplasms in the Egyptian Toad by gibberellin A3. Oncology; 45: 61–64.
- El-Mofty MM, Sakr SA, Rizk AM, Moussa EA (1994). Carcinogenic effect of gibberellin A3 in Swiss Albino mice. Nutr cancer; 21: 183-190.
- Folkes LK, Dennis MF, Stratford MRL, Candeias LP, Wardman P (1999). Peroxidase-catalyzed effects of indole-3-acetic acid and analogues on lipid membranes, DNA, and mammalian cells in vitro. Biochem Pharmacol.; 57: 375–382.
- Frukawa S, Masayoshi A, Usuda K. (2004). Indole-3-acetic acid induces microencephaly in rat fetuses. Toxicologic Pathology; 32: 659-667.
- Hall BK, Miyake T (1992). The membranous skeleton: the role of cell condensations in vertebrate skeletogenesis. Anat Embryol Berl.; 186(2): 107-24.
- Hall BK, Miyake T (1995). Divide, accumulate, differentiate: cell condensation in skeletal development revisited.Int J Dev Biol.; 39(6): 881-93.
- Hsiao G, Shen MY, Lin KH, Chou CY, Tzu NH, Lin CH, Chou DS, Chen TF, Sheu JR (2004). Inhibitory activity of kinetin on free radical formation of activated platelets in vitro and on thrombus formation in vivo. Eur J Pharmacol.; 465: 281–87.
- Hu T, Dryhurst G (1997). Eletrochemical and peroxidase O2-mediated oxidation of indole-3-acetic acid at physiological pH. J Electroanal Chem.; 432: 7–18.
- Hussein WF, Farahat FV, Abass MA, Shehata AS (2011). Hepatotoxic Potential of Gibberellic Acid (GA3) in Adult Male Albino Rats. Life Sci J.; 8(3):373-383.

- Kochupillai N (2008). The physiology of vitamin D: Current concepts. Indian J Med Res.; 127: 256-262
- Lehmann B, Meurer M. (2010). Vitamin D metabolism. Dermatol Ther.; 23(1): 2-12.
- Mickel LG (1978). "Plant Growth Regulators" Controlling biological behavior with chemicals, Chem Eng News; 56: 18.
- Mickey DD, Paulson DF (1978). Prostate and transitional cell carcinoma: radioimmunoassay of viral tumor-associated antigens. Natl Cancer Inst Monogr.; 49: 51-3
- Ozmen M, Topcuoglu SF, Bozcuk S, Bozcuk NA (1995). Effects of abscisic acid and gibberellic acid on sexual differentiation and some physiological parameters of laboratory mice, Turk J Biol.; 19: 357–364.
- Qureshi GA, Baig SM (1993). The role of tryptophan, 5-hydroxy-indole-3-acetic acid and their protein-binding in uremic patients. Biochemistry and Molecular Biology International; 29: 411 –419.
- Rogerson R, Gallagher ML, Zallen EM, Mc Mi1len BA (1991). Urinary tryptophan-metabolites in diabetic and non diabetic juveniles. Nutrition Research; 11: 1251–1256.
- Sakr SA, Okdah YA, El-Abd SF (2003). Gibberellin A3 induced histological and histochemical alterations in the liver of albino rats. Science Asia; 29: 327-331.
- Serrani JC, Fos M, Atarés A, García-Martínez JL (2007a). Effect of gibberellin and auxin on parthenocarpic fruit growth induction in the cv Micro-Tom of tomato. J Plant Growth Regul.; 26: 211-221.
- Serrani JC, Sanjuan R, Ruiz-Rivero O, Fos M, García-Martínez JL (2007b). Gibberellin Regulation of Fruit-Set and Growth in Tomato. Plant Physiol.; 145: 246-257.
- Soliman HAE, Mantawy MM, Hassan HM (2010). Biochemical and Molecular Profiles of Gibberellic Acid Exposed Albino Rats. J Am. Science; 6: 224-229.
- Troudi A, Bouaziz H, Soudani N, Ben Amara I, Boudawara T, Touzani H, Lyoussi B, Zeghal N (2011). Neurotoxicity and oxidative stress induced by gibberellic acid in rats during late pregnancy and early postnatal periods: Biochemical and histological changes. Experimental and Toxicologic Pathology.
- Troudi A, Samet AM, Zeghal N (2010). Hepatotoxicity induced by gibberellic acid in adult rats and their progeny. Exp Toxicol Pathol.; 62(6):637-642.
- Tulice Y, Celik I (2006). Influence of subacute and subchronic treatment of abcisic acid and gibberellic acid on serum marker enzymes and erythrocyte and tissue antioxidant defense systems and lipidperoxidation in rats. Pestic Biochem Phys.; 86: 85–92.
- Ustun H, Tecimer T, Ozmen M, Topcuoglu SF, Bozcuk NA (1992). Effects of gibberellic acid and benzoprenin on mice. Histopatalogic review, Ank Patol. Bu" lt.; 9: 36–40.
- Vessey DA (1996). Metabolism of xenobiotics by the human liver. Zakin D Boyer TD eds. Hepatology: a textbook of liver disease 3rd ed. Saunders Philadelphia; WB 257-305.

Visscher NS (1983). Special report dietary plant growth hormones affects insect growth and reproduction, Bull. Plant Growth Reg Soc Am.; 11:

WHO (1990). Public health impact of pesticides used in agriculture. WHO/UNEP, Geneva.

إيضاح التأثير السام والمسبب للتشوهات لبعض الهرمونات النباتية على أجنة الجرذان البيضاء

حسن إبراهيم الصياد، محمد حسن أبوعجلة، # محمد محمد مرتضى، منال محمد رمضان و محمد عزت البيه

قسم علم الحيوان ، كلية العلوم ، جامعة المنصورة # معهد بحوث وقاية النبات ، وزارة الزراعة

تهدف الدراسة إلى إيضاح التأثير السام للهرمونات النباتية والتى تستخدم بصفة واسعة فى الزراعة. أحد هذه الهرمونات النباتية هو (حامض الجبرياليك) والثانى هو (الإندول حامض الخليك)؛ ولقد تم إعطاء كلا من اللهرمونين النباتيين إلى الجرذان بنسبة 200 جزء فى المليون لكل ٢٠٠ محلول ملح يوم بعد يوم ولمدة أربعين يوم قبل الحمل وحتى اليوم الرابع عشر أو السادس عشر أو الولادة.

وتتلخص نتائج تلك الدراسة في الأتي :-

التأثير على الحمل والأجنة والولائد:

أظهرت الأمهات المعاملة بالهرمونات النباتية زيادة ملحوظه من حالات الاجهاض . ولقد تم تسجيل زيادة ملحوظة في الأمهات المعاملة مقارنة بالمجموعة الضابطة ولقد صاحب ذلك العديد من التشوهات الخلقية سواء تقوس الظهر وتشوه الذيل ووجود تجمعات دموية الى غير ذلك من تشوهات الأطراف. كما لوحظ تقلص وزن الجسم في أجنة الأمهات المعاملة بالهرمونات النباتية. كما لوحظ از دياد معدل امتصاص الأجنة في أجنة الأمهات المعاملة مقارنة بالمجموعة الضابطة.

أظهرت ولائد الأمهات المعاملة بكلا اللهرمونات النباتية بعد إنتهاء المعاملة بعض التشوهات الخلقية متمثلة في نقص تكون كل من الطرفين الأمامي والخلفي، اختزال ملحوظ لوزن الجسم وطوله، بالإضافة إلى ما سبق فلقد أظهرت المعاملة تقلص عظام الهيكل متمثلة في اللحي - اللوح -الحوض - الطرف الأمامي والخلفي .

التأثير على مراكز التعظم:

كما سجلت الدراسة أن أجنة الأمهات المعاملة بالهرمونات النباتية ظهور تخلف ملحوظ في مراكز التعظم في المناطق المحورية في القوس التعظم في المناطق المحورية في القوس اللامي، سندان ، المطرقة ، ومنطقة الطبلة و الركاب . بالاضافة الى ذلك تم الكشف عن عدد الحالات المصحوبه بتشوه اليافوخ الأمامية والخلفية في الأجنة لأمهات المعاملة بالهرمونات النباتية . كما سجلت الدراسة تقلص مراكز التعظم في منطفة الفك السفلي وعظام الطرف الأمامي والخلفي.

قام بتحكيم البحث

كلية الزراعة – جامعة المنصورة كلية العلوم – جامعة المنصورة أد / على على عبد الهادى أد / سعاد احمد خليفه