



UVC effect on growth, structure and bio-contents of *Foeniculum vulgare*

Hager Yuns¹, Abu Bakr El-Bediwi^{*1}, Hamed M El-Shora²

¹Physics Department, Faculty of Science, Mansoura University, Egypt

²Botany Department, Faculty of Science, Mansoura University, Egypt

baker_elbediwi@yahoo.com

Received: 6/6/2021
Accepted: 4/7/2021

Abstract: Morphological structures (growth, shape and weight), internal structure (order and position of chemical compound) and bio-contents (enzymes, vitamins and antioxidants for normal and irradiated *Foeniculum vulgare*) are studied. The growth of *Foeniculum vulgare* seeds are effected after exposure to UVC. The weight of *Foeniculum vulgare* seeds also decreased by exposure to UVC. Continuous increase in the *Foeniculum vulgare* enzyme activity, tocopherol content, vitamin C, total phenol content and antioxidant activity was observed by increasing the exposure time up to 3 hours then declined in the fourth hour.

keywords: *Foeniculum vulgare*, UVC, Molecular structure, Antioxidant activity, Phenol, Vitamins

1. Introduction

Foeniculum vulgare is a flowering plant species in the carrot family. It contains 6.3% of moisture, 9.5% protein, 10% fat, 13.4% minerals, 18.5% fibre and 42.3% carbohydrates. Ultraviolet (UV) constituted approximately 8- 9 % of total solar radiation and considers a part of non-ionizing rays of the electromagnetic spectrum region [1, 2]. Plants sensitive to UV can respond through accumulating some compounds in their outer tissue layers, which protect sensitive parts from damage by UV. The main enzymes involved in biosynthesis of these compounds are induced by UV irradiation via gene activation [3]. Higher UV radiation can inhibit plant growth and development of plants as well as reproduction and suppress photosynthesis [4, 5]. UV in the range 2800- 4000 Å as a small fraction of the solar radiation reaching the terrestrial ecosystems is an important modulator of plant physiology [6]. The physiological, biochemical processes, leaf chlorophyll, protein content, and peroxidase enzyme activity in plants can be affected by UVC [7, 8]. The effect of UVC on the germination percentage, germination rate, radicle length, and plumule length of maize and sugar beet seeds has been reported [9]. Also UVC increased the germination percentage of the groundnut [10]. Nonetheless that UVC radiation did not

significantly affect the germination percentage of *Acacia ampliceps* [11]. The effect of UVC on growth, internal structure, free radical and enzymes of *Nigella Sativa* have been studied [12]. Furthermore, UVC effect on morphological characteristics and the content in *garden cress* studied and analyzed [13]. Internal structure and contents of *Ammi majus* changed by exposure to UVC for different time intervals and different distances [14]. The objective of the present research was to study the effect of ultra violet- C on morphological structure and medical contents such as enzymes, vitamins and antioxidant activity for *Foeniculum vulgare*.

The aim of this study is to investigate the morphological structures (growth, shape and weight), internal structure (order and position of chemical compound) and bio-contents (enzymes, vitamins and antioxidants for normal and irradiated *Foeniculum vulgare*).

Materials and experimental techniques

Seeds of *Foeniculum vulgare* are received from Egyptian ministry of agriculture. The irradiated used system consists of fluorescent lamp (Type-C with λ from 2000-2800 Å), its power equal to 15 watt. Also the system covered totally with aluminum foil to illuminate the sample from all sides. Internal structure of *Foeniculum vulgare* was studied by

molecular structure such as C- C bond at 1649 cm^{-1} , C-H bond at 2924 cm^{-1} and O- H bond at $\sim 3421\text{ cm}^{-1}$ bonded of *Foeniculum vulgare* changed after exposure to UVC. These results reveled that these molecules absorb energy from UV which caused vibrated or break bonds linked it, produced its position, intensity and broad nesses change. These results agree with our previous studies [12- 14].

Table 1:- x-ray data for *Foeniculum vulgare*

	d Å	2θ	Intensity	Area	FWHM
Normal	4.10865	21.612	32.1	6.1	0.572
	3.11758	28.610	21.3	4.9	0.414
1 hr. at 5 cm	4.11939	21.555	34.7	356.8	11.21
	3.13292	28.467	28.5	2.4	0.187
2 hrs. at 5 cm	4.12158	21.543	33.2	125.8	6.4°
	3.13970	28.404	22.8	2.6	0.29
4 hrs. at 5 cm	4.12158	21.543	36.1	193.95	8.196
	3.13970	28.404	25.9	4.30	0.24

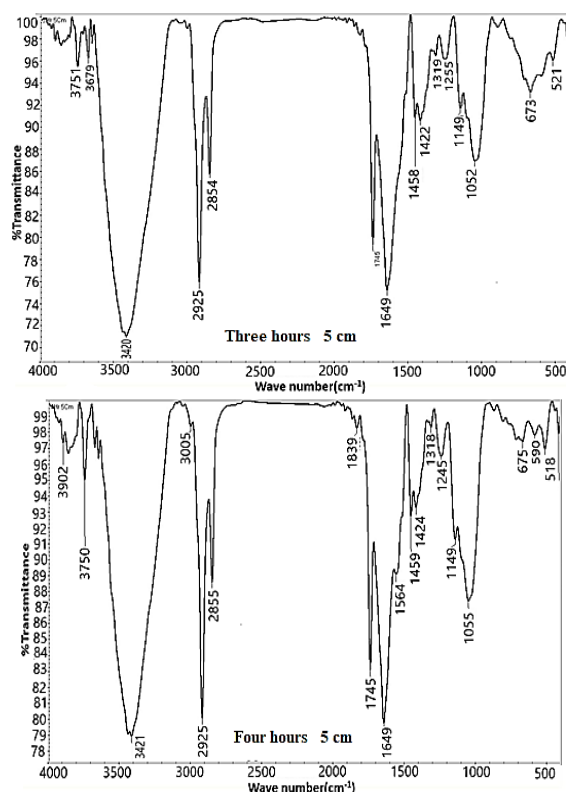
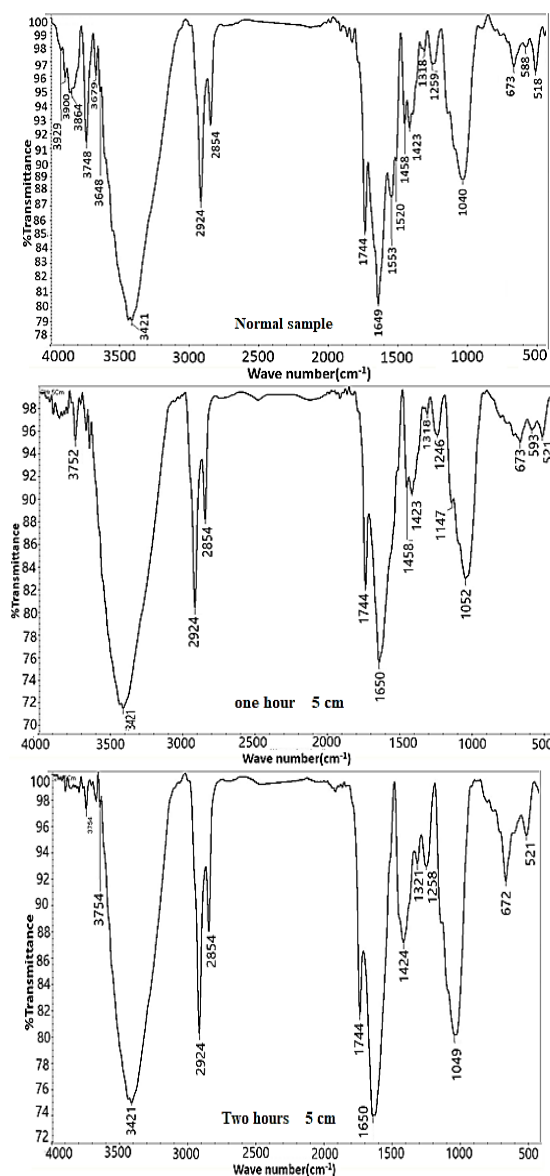


Figure 2: IR spectra for *Foeniculum vulgare*

Table 2: IR spectrum analysis of *Foeniculum vulgare*.

Exposure Time/ at 5 cm	Position cm^{-1}	%Transmittance
Normal	1052	82.93
	1649	80.15
	2925	87.28
One hour	1052	82.93
	1650	75.5
	2924	80.38
Two hours	1049	80.08
	1650	73.91
	2924	80.2
Four hours	1055	87.37
	1649	79.7
	2925	80.03
	3421	78.88



Antioxidant activity of methanol extract from *Foeniculum vulgare*

The antioxidant activity of methanolic and ethanol extracts from *Foeniculum vulgare* exposed to UVC at 5 cm for 1, 2, 3 and 4 hours presented in Figure 3 indicated continuous increase in the antioxidant activity throughout the four hours of exposure at each individual concentration of ($20, 40, 60, 80$ and $100\text{ }\mu\text{g ml}^{-1}$). Also, it was found that, the increase in the antioxidant activity was continuous until the third hour, then decreased after the fourth hour.

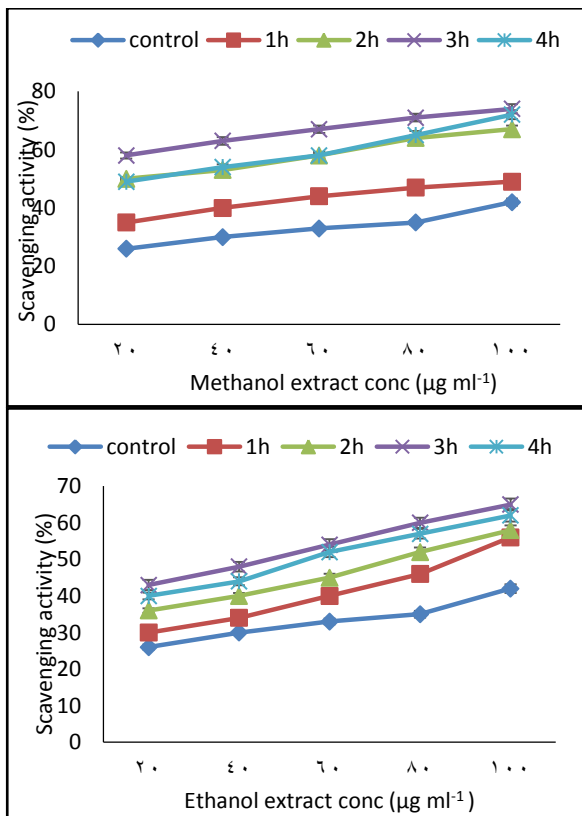


Figure 3: antioxidant activity of methanol and ethanol extracts from *Foeniculum vulgare*

Total phenol content of methanol extract from *Foeniculum vulgare*

Figure 4 shows the continuous increase in total phenol content in the methanol extract throughout the first three hours. However, after the fourth hour the total phenol content declined. The total phenol content value of the ethanol extract from *Foeniculum vulgare* exposed to UVC at 5 cm also increased in the same manner recorded for methanol extract.

Vitamin C content of ethanol and methanol extract from *Foeniculum vulgare*

Vitamin C content of methanol and ethanol extracts from *Foeniculum vulgare* illustrated in Figure 5 indicated continuous increase in vitamin C in *Foeniculum vulgare* by exposure to UVC at 5 cm for 1, 2 and 3 hours but after the fourth hour the vitamin C content decreased.

Tocopherol content of methanol and ethanol extracts from *Foeniculum vulgare*

Figure 6 shows the tocopherol contents in methanol and ethanol extracts from *Foeniculum vulgare* increased by exposed to UVC at 5 cm and the exposure intervals 1h, 2h, and 3h whereas exposure for 4 h resulted in reduction of tocopherol.

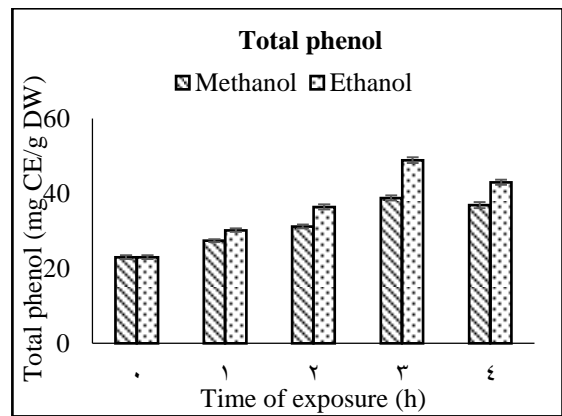


Figure 4: Total phenol content of methanol and ethanol extracts from *Foeniculum vulgare*

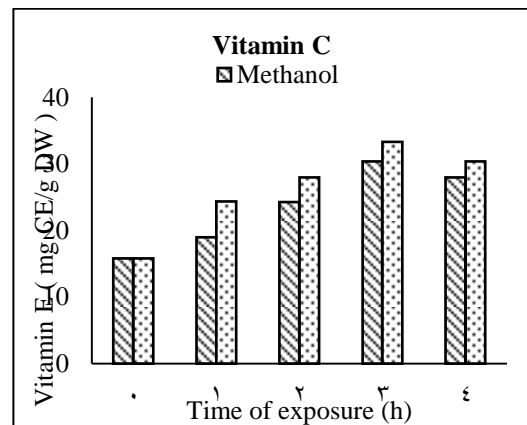


Figure 5: Vitamin C content of methanol and ethanol extracts from *Foeniculum vulgare*

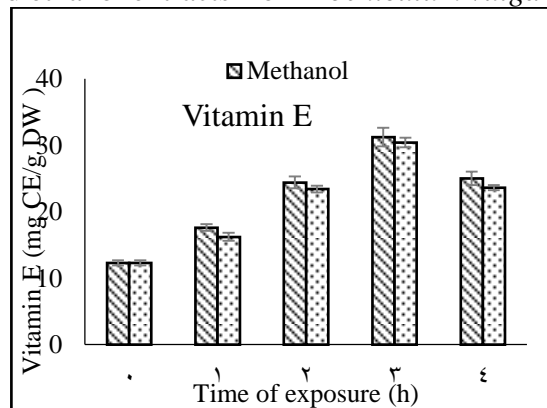


Figure 6: Vitamin E content of methanol and ethanol extracts from *Foeniculum vulgare*

Antioxidant enzymes in *Foeniculum vulgare*
Superoxide dismutase (SOD)

The activity of SOD was estimated in extract prepared from *Foeniculum vulgare* and the results shown in Figure 7 indicate continuous increase in the enzyme activities after exposure to UVC for 1, 2 and 3 hours after which it declined by exposure for 4 hours.

Ascorbate peroxidase (APX)

The APX activity was measured in *Foeniculum vulgare* after exposure to UVC for

1, 2, 3 and 4 hours and the results presented in Figure 7 revealed an increase in the enzyme activity by exposure to UVC until the third hour but after the fourth hour the activity declined.

Glutathione reductase (GR)

The activity of glutathione reductase in extract prepared from *Foeniculum vulgare* was measured and shown in Figure 7. The result indicated continuous increase in the enzyme activity by increasing the exposure time to 3 hours then declined during the fourth hour.

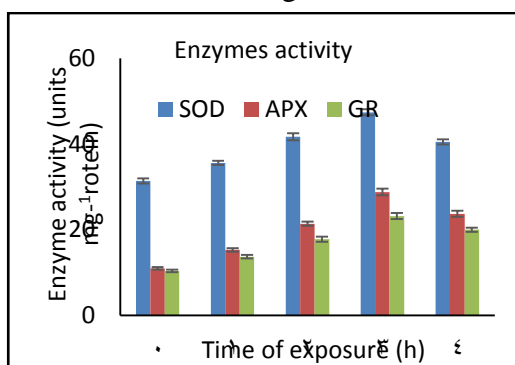


Figure 7: Enzymes activity in *Foeniculum vulgare*

Seeds growth (morphological structure)

Figure 8 shows *Foeniculum vulgare* seeds growth before and after exposure to UVC for different interval times (1, 2, 3 and 4 hours) at distance 5 cm. The growth such as plant shape, straightness of branch growth, greenness, and number of grown seeds was affected by exposure to UVC. It means that UVC effected on the internal structure of seeds as proved before in x-ray and IR analysis.



Figure 8: seeds growth before and after exposure to UVC

Seeds weight (volumetrically change)

The weight of *Foeniculum vulgare* seeds decreased after exposure to UVC for time intervals (1, 2, 3 and 4 hours) at distance 5 cm as seen in Table 3. It means that UVC effected the seed's geometrical structure

Table 3: seeds weight (g) before and after exposure to UVC

Time	Plant seeds weight (g)
Normal	5
1hour	4.85
2hour	4.84
3hour	4.79
4hour	4.77

Conclusion

The growth criteria of *Foeniculum vulgare* affected after exposure to UVC. Thus, UVC radiation effected on the internal structure of seeds as proved before by x-ray and IR analysis. The seeds weight decreased by exposure to UVC. It means UVC effected the geometrical structure of seeds. *Foeniculum vulgare* enzyme activity, tocopherol content, vitamin C, total phenol content and antioxidant activity values increased by increasing the exposure time to 3 hours then declined during the fourth hour. The increase of these contents during the first three hours of exposure may be due to induction of the enzymes involved in biosynthesis of these components. However, this decline may be due to inactivation of these enzymes at prolonged time of UVC exposure.

4. References

- Hollósy, F. (2002). Effects of ultraviolet radiation on plant cells. *Micron*, **33**(2), 179-197.
- Frederick, J. E. (1993). Ultraviolet sunlight reaching the earth's surface: a review of recent research. *Photochemistry and Photobiology*, **57**(1), 175-178.
- Teramura, A. H., Sullivan, J. H., Abrol, Y. P., Wattal, P. W., Ort, D. R., & Gnanam, A. (1991). Field studies of UV-B radiation effects on plants: Case histories of soybean and loblolly pine. *Abrol, YP, PW Wattal, DR Ort and A. Gnanam (Eds.)*, 147-161.
- Hopkins, L., Bond, M. A., & Tobin, A. K. (2002). Ultraviolet- B radiation reduces the rates of cell division and elongation in the primary leaf of wheat (*Triticum aestivum* L. Cv Maris Huntsman). *Plant, Cell & Environment*, **25**(5), 617-624.

5. Jansen, M. A., Gaba, V., & Greenberg, B. M. (1998). Higher plants and UV-B radiation: balancing damage, repair and acclimation. *Trends in Plant Science*, **3(4)**, 131-135.
6. Paul, N. D. & Gwynn-Jones, D. (2003). Ecological roles of solar UV radiation: towards an integrated approach. *Trends in Ecology & Evolution*, **18(1)**, 48-55.
7. Salama, H. M., Al Watban, A. A., & Al-Fughom, A. T. (2011). Effect of ultraviolet radiation on chlorophyll, carotenoid, protein and proline contents of some annual desert plants. *Saudi Journal of Biological Sciences*, **18(1)**, 79-86.
8. Falconí, C. E., & Yáñez- Mendizábal, V. (2018). Efficacy of UV- C radiation to reduce seedborne anthracnose (*Colletotrichum acutatum*) from Andean lupin (*Lupinus mutabilis*). *Plant Pathology*, **67(4)**, 831-838
9. Sadeghianfar, P., Nazari, M., & Backes, G. (2019). Exposure to ultraviolet (UV-C) radiation increases germination rate of maize (*Zea maize L.*) and sugar beet (*Beta vulgaris*) seeds. *Plants*, **8(2)**, 49.
10. Neelamegam, R., & Sutha, T. (2015). UV-C irradiation effect on seed germination, seedling growth and productivity of groundnut (*Arachis hypogaea L.*). *International Journal of Current Microbiology and Applied Sciences*, **4(8)**, 430-443.
11. Shetta, D. N., & Areaf, M. I. (2009). Impact of ultraviolet-C radiation on seed germination and chlorophyll concentration of some woody trees grown in Saudi Arabia. *J. Agric. Environ. Sci*, **8**, 1-21.
12. El-Bediwi, A. B., Hasanin, S., Abdelrazek, A., & El-Shora, H. M. (2018). Effect of ultraviolet on morphological and secondary metabolites content of garden cress. *International Journal of Scientific Research in Science, Engineering and Technology*, **4(1)**, 187- 194.
13. El-Bediwi, A. B., Hasanin, S., & Abdelrazek, A. (2018). Influence of UVC on growth behavior. *Internal Structure, Enzymes and Free Radical of Nigella Sativa Plant*, **13(2)**, 142.
14. A.B. El-Bediwi, H. Yuns., & H. M El-Shora, (2020), UVC Radiation Effects on the internal structure and medical contents of *Ammi majus*. *Int J Biotech & Bioeng*. **6:1** 07-21.
15. Hodges, D. M., Andrews, C. J., Johnson, D. A., & Hamilton, R. I. (1996). Antioxidant compound responses to chilling stress in differentially sensitive inbred maize lines. *Physiologia Plantarum*, **98(4)**, 685-692.
16. Hira, T., Hara, H., & Tomita, F. (2001). Theory and practice on enzymes and other proteins, 1994. Bioscience, *Biotechnology, and Biochemistry*, **65(5)**, 1007-1015.
17. Singleton, V. L., & Rossi, J. A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*, **16(3)**, 144-158.
18. Dhindsa, R. S., & Matowe, W. (1981). Drought tolerance in two mosses: correlated with enzymatic defence against lipid peroxidation. *Journal of Experimental Botany*, **32(1)**, 79-91.
19. Nakano, Y., & Asada, K. (1987). Purification of ascorbate peroxidase in spinach chloroplasts; its inactivation in ascorbate-depleted medium and reactivation by monodehydroascorbate radical. *Plant and Cell Physiology*, **28(1)**, 131-140.
20. Goldberg, D. M., & Spooner, R. J. (1983). Methods of enzymatic analysis. *Bergmeyer HV*, **3**, 258-265.